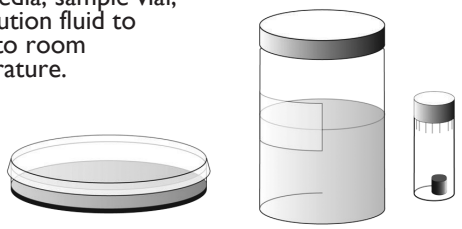
Urine & Blood Sample Preparation Illustrations

Instructions for preparing Urine Colony Count, Identification, and Susceptibility Samples

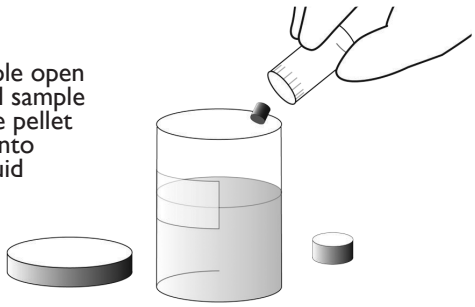
1.

Before testing allow the agar media, sample vial, and dilution fluid to warm to room temperature.



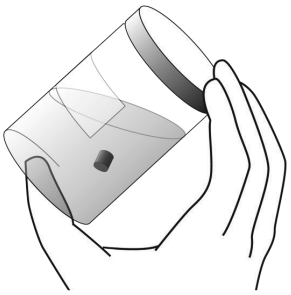
2.

For each sample open the lyophilized sample vial. Empty the pellet from the vial into the dilution fluid container.



3.

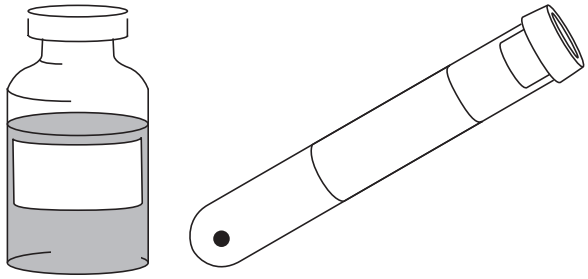
Reseal the dilution fluid container, then mix the contents, allow pellet to dissolve for 20 minutes and mix again. No incubation is required before use.



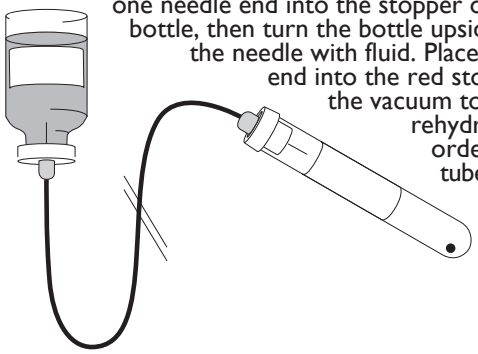
4. The entire contents of the container (which now contains the dissolved lyophilized pellet) simulates a urine sample. Mix and immediately proceed to test as you would a patient sample in your laboratory.

Instructions for preparing Blood Culture Samples

1. Allow samples to come to room temperature. Do not remove the red stopper from the vacutainer (this tube contains vacuum). Lift the aluminum tab on the rehydration bottle and expose the rubber septum top. Disinfect the septum top on the rehydration bottle and the red stopper vacutainer tube.

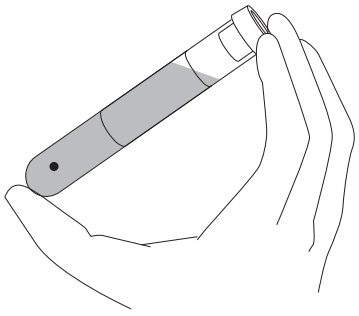


2. **Closed System Method:** Using aseptic technique, remove 10 mls of rehydration fluid from the rehydration bottle with a butterfly set-up. Transfer the rehydration fluid into the red stopper vacutainer tube by placing one needle end into the stopper of the rehydration bottle, then turn the bottle upside down to cover the needle with fluid. Place the other needle end into the red stopper tube. Allow the vacuum to fill the tube with rehydration fluid. (If the order is reversed, the tube will lose vacuum. If vacuum is lost, proceed to Open System Method as described in Testing Instructions.)



3.

Mix sample thoroughly until pellet is dissolved.



4.

Aseptically, inoculate half of the sample/ rehydration fluid suspension (5.0 mls) into each blood culture bottle using the method of your choice (a sterile needle and syringe, vacutainer, or butterfly set up). Incubate and process inoculated blood culture bottle(s) following your laboratory's procedure.